

## Two New Diterpenes from *Solidago canadensis*

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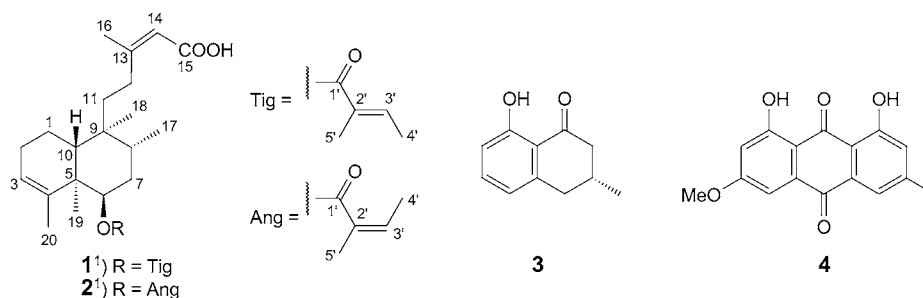
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Two new clerodane-type diterpenes, solidagocanins A and B (**1** and **2**, resp.), as well as two known compounds, mellein (**3**) and physcion (**4**), were isolated from the aerial parts of *Solidago canadensis*. The structures of the new compounds were elucidated by spectroscopic analyses, including 2D-NMR techniques.

**Introduction.** – Diterpenes and polyacetylenes are common constituents in members of the tribe Astereae [1–5]. In recent years, a large number of reports have appeared dealing with the biological activities of these compounds. Diterpenes exhibit insect-antifeedant, antifungal, antibacterial, and antiviral activities [3][6]. In the course of searching for the new bioactive natural products from the Asteraceae plants, two new clerodane-type diterpenoids, solidagocanins A and B<sup>2)</sup> (**1** and **2**, resp.), as well as two known compounds, mellein (**3**) and physcion (**4**), were isolated from the aerial parts of *Solidago canadensis*. This article describes the isolation and structure elucidation of the new compounds.



**Results and Discussion.** – Repeated column chromatography of the AcOEt extract from the aerial parts of *S. canadensis* yielded compounds **1–4**.

<sup>1)</sup> These two authors have contributed equally to this work.

<sup>2)</sup> Trivial atom numbering; for systematic names, see *Exper. Part*.

Solidagocanin A<sup>2</sup>) (**1**) was obtained as a white powder and assigned to possess a molecular formula C<sub>25</sub>H<sub>38</sub>O<sub>4</sub> on the basis of the HR-ESI-MS (*m/z* 403.2835 ([*M* + H]<sup>+</sup>)), which indicated seven degrees of unsaturation. The <sup>1</sup>H-NMR spectrum (Table 1) showed two secondary Me groups ( $\delta$ (H) 0.85 (*d*, *J* = 6.4 Hz) and 1.78 (*d*, *J* = 7.0 Hz)) and five tertiary Me groups ( $\delta$ (H) 1.05, 1.25, 1.54, 1.79, and 2.20 (*s*)). As determined by a DEPT experiment, the down-field <sup>13</sup>C-NMR signals (Table 2) corresponded to two C=O groups ( $\delta$ (C) 172.3 and 167.6), six olefinic C-atoms ( $\delta$ (C) 115.0, 124.2, 129.1, 136.8, 138.2, and 164.3), and one O-bearing C-atom ( $\delta$ (C) 74.4); the high-field region showed seven Me ( $\delta$ (C) 12.2, 14.4, 15.4, 18.3, 19.5, 24.7, and 28.6), five CH<sub>2</sub> ( $\delta$ (C) 21.8, 26.5, 31.5, 34.2, and 35.6), and two CH groups ( $\delta$ (C) 31.9 and 44.4) as well as two quaternary C-atoms ( $\delta$ (C) 38.5 and 42.9) (Table 2). The <sup>1</sup>H-NMR signals at  $\delta$ (H) 6.79 (*m*, H–C(3')), 1.78 (*d*, *J* = 7.0 Hz Me(4')), and 1.79 (*s*, Me(5')), along with the corresponding <sup>13</sup>C-NMR signals at  $\delta$ (C) 167.6 (C(1')), 129.1 (C(2')), 136.8 (C(3')), 14.4 (C(4')), and 12.2 (C(5')) indicated the presence of a tigloyl (Tig) group [4]. The <sup>1</sup>H-NMR, <sup>13</sup>C-NMR, and HMBC data (Fig.) of **1** suggested that **1** had the same constitution as (6 $\beta$ )-6-(tigloyloxy)kolavenic acid [4]. Comparison of the <sup>13</sup>C-NMR spectra of **1** and (6 $\beta$ )-6-(tigloyloxy)kolavenic acid (Table 2) showed that they only differed in the configuration of the C=C bond at C(13). The configuration of this C=C bond was determined by NOESY experiments. H–C(14) and the Me(16) were *trans*-oriented in (6 $\beta$ )-6-(tigloyloxy)kolavenic acid [4]. However, the NOESY correlation between Me(16) and H–C(14) of **1** indicated that H–C(14) and the Me(16) were *cis*-oriented (Fig.), similar to the situation in (7 $\alpha$ ,13*Z*)-7-(acetyloxy)kolavenic acid [4]. Thus, the structure of **1** was elucidated as shown in the Figure, and it was named solidagocanin A.

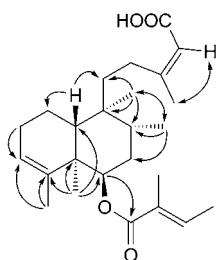
Table 1. <sup>1</sup>H-NMR Data (at 27°, CDCl<sub>3</sub>, 400 MHz) of **1** and **2**<sup>2</sup>).  $\delta$  in ppm, *J* in Hz.

H-Atom	<b>1</b>	<b>2</b>
CH <sub>2</sub> (1)	1.33–1.42, 1.94–2.03 ( <i>2m</i> )	1.31–1.41, 1.90–2.00 ( <i>2m</i> )
CH <sub>2</sub> (2)	1.89–2.08 ( <i>m</i> )	1.83–2.02 ( <i>m</i> )
H–C(3)	5.49 ( <i>br. s</i> )	5.47 ( <i>br. s</i> )
H–C(6)	4.98 ( <i>br. s</i> )	5.01 ( <i>br. s</i> )
CH <sub>2</sub> (7)	1.55–1.63, 1.75–1.84 ( <i>2m</i> )	1.50–1.60, 1.75–1.85 ( <i>2m</i> )
H–C(8)	1.88–1.96 ( <i>m</i> )	1.83–1.91 ( <i>m</i> )
H–C(10)	1.51–1.58 ( <i>m</i> )	1.46–1.54 ( <i>m</i> )
CH <sub>2</sub> (11)	1.63–1.70, 1.82–1.90 ( <i>2m</i> )	1.58–1.66, 1.77–1.86 ( <i>2m</i> )
CH <sub>2</sub> (12)	2.02–2.12, 2.22–2.32 ( <i>2m</i> )	2.00–2.10, 2.20–2.30 ( <i>2m</i> )
H–C(14)	5.71 ( <i>s</i> )	5.69 ( <i>s</i> )
Me(16)	2.20 ( <i>s</i> )	2.18 ( <i>s</i> )
Me(17)	0.85 ( <i>d</i> , <i>J</i> = 6.4, 3 H)	0.83 ( <i>d</i> , <i>J</i> = 6.4)
Me(18)	1.05 ( <i>s</i> )	1.02 ( <i>s</i> )
Me(19)	1.25 ( <i>s</i> )	1.24 ( <i>s</i> )
Me(20)	1.54 ( <i>s</i> )	1.54 ( <i>s</i> )
H–C(3')	6.79 ( <i>m</i> )	5.97 ( <i>m</i> )
Me(4')	1.78 ( <i>d</i> , <i>J</i> = 7.0)	1.96 ( <i>d</i> , <i>J</i> = 7.0)
Me(5')	1.79 ( <i>s</i> )	1.81 ( <i>s</i> )

Table 2.  $^{13}\text{C}$ -NMR Data of **1**, **2**, and (13Z)-**1** [4]<sup>a</sup>). Recorded at 27°, CDCl<sub>3</sub>, 100 MHz;  $\delta$  in ppm.

C-Atom	<b>1</b>	(13Z)- <b>1</b>	<b>2</b>
C(1)	34.2 ( <i>t</i> )	34.6 ( <i>t</i> )	34.2 ( <i>t</i> )
C(2)	26.5 ( <i>t</i> )	26.5 ( <i>t</i> )	26.5 ( <i>t</i> )
C(3)	124.2 ( <i>d</i> )	124.2 ( <i>d</i> )	124.5 ( <i>d</i> )
C(4)	138.2 ( <i>s</i> )	138.2 ( <i>s</i> )	138.0 ( <i>s</i> )
C(5)	42.9 ( <i>s</i> )	42.9 ( <i>s</i> )	42.9 ( <i>s</i> )
C(6)	74.4 ( <i>d</i> )	74.4 ( <i>d</i> )	74.0 ( <i>d</i> )
C(7)	31.5 ( <i>t</i> )	31.5 ( <i>t</i> )	31.6 ( <i>t</i> )
C(8)	31.9 ( <i>d</i> )	31.9 ( <i>d</i> )	31.9 ( <i>d</i> )
C(9)	38.5 ( <i>s</i> )	38.5 ( <i>s</i> )	38.4 ( <i>s</i> )
C(10)	44.4 ( <i>d</i> )	44.4 ( <i>d</i> )	44.3 ( <i>d</i> )
C(11)	21.8 ( <i>t</i> )	21.8 ( <i>t</i> )	21.6 ( <i>t</i> )
C(12)	35.6 ( <i>t</i> )	35.5 ( <i>t</i> )	35.6 ( <i>t</i> )
C(13)	164.3 ( <i>s</i> )	162.9 ( <i>s</i> )	164.2 ( <i>s</i> )
C(14)	115.0 ( <i>d</i> )	116.2 ( <i>d</i> )	115.0 ( <i>d</i> )
C(15)	172.3 ( <i>s</i> )	172.2 ( <i>s</i> )	172.0 ( <i>s</i> )
C(16)	19.5 ( <i>q</i> )	19.4 ( <i>q</i> )	19.5 ( <i>q</i> )
C(17)	15.4 ( <i>q</i> )	15.3 ( <i>q</i> )	15.4 ( <i>q</i> )
C(18)	28.6 ( <i>q</i> )	28.5 ( <i>q</i> )	28.6 ( <i>q</i> )
C(19)	24.7 ( <i>q</i> )	24.7 ( <i>q</i> )	24.7 ( <i>q</i> )
C(20)	18.3 ( <i>q</i> )	18.2 ( <i>q</i> )	18.3 ( <i>q</i> )
C(1')	167.6 ( <i>s</i> )	167.5 ( <i>s</i> )	167.1 ( <i>s</i> )
C(2')	129.1 ( <i>s</i> )	129.1 ( <i>s</i> )	128.3 ( <i>s</i> )
C(3')	136.8 ( <i>d</i> )	136.7 ( <i>d</i> )	137.7 ( <i>d</i> )
C(4')	14.4 ( <i>q</i> )	14.3 ( <i>q</i> )	15.5 ( <i>q</i> )
C(5')	12.2 ( <i>q</i> )	12.1 ( <i>q</i> )	20.7 ( <i>q</i> )

<sup>a</sup>) The known compound (13Z)-**1** was named (6 $\beta$ )-6-(tigloyloxy)kolavenic acid (= (2*E*)-3-methyl-5-[(1*S*,2*R*,4*R*,4*aR*,8*aR*)-1,2,3,4,4*a*,7,8,8*a*-octahydro-1,2,4*a*,5-tetramethyl-4-[(2*E*)-2-methyl-1-oxobut-2-en-1-yl]oxy]naphthalen-1-yl]pent-2-enoic acid).

Figure. Key HMBC (H → C) and NOESY (H ↔ H) correlations in **1**

Solidagocanin B<sup>2</sup>) (**2**), obtained as a white powder, had a molecular formula C<sub>25</sub>H<sub>38</sub>O<sub>4</sub> according to the HR-ESI-MS (*m/z* 403.2832 ([*M* + H]<sup>+</sup>)). The NMR, IR, and UV of **2** were quite similar to those of **1**, except for the characteristic signals due to an angeloyl (= (2*Z*)-2-methyl-1-oxobut-2-en-1-yl; Ang) group instead of the tigloyl (= (2*E*)-2-methyl-1-oxobut-2-en-1-yl; Tig) group in **1**. The <sup>1</sup>H-NMR signals of **2** at  $\delta$ (H) 5.97 (*m*, H–C(3')), 1.96 (*d*, *J* = 7.0 Hz, Me(4')), and 1.81 (*s*, Me(5')), along with

the corresponding  $^{13}\text{C}$ -NMR signals at  $\delta(\text{C})$  167.1 (C(1')), 128.3 (C(2')), 137.7 (C(3')), 15.5 (C(4')) and 20.7 (C(5')) indicated the presence of an Ang group [4][7]. The HMBC between H–C(6) at  $\delta(\text{H})$  5.01 and C(1') at  $\delta(\text{C})$  167.1 suggested that the Ang group was located at C(6). The NOESY correlation between Me(16) and H–C(14) of **2** indicated that H–C(14) and Me(16) were *cis*-oriented as in **1**, but different from the configuration of (6 $\beta$ )-6-(angeloyloxy)kolavenic acid [4].

The known compounds were identified as mellein (**3**) [8] and physcion (**4**) [9] by comparing their IR, UV, and NMR data with those reported in the literature.

### Experimental Part

*General.* Column chromatography (CC): silica gel ( $\text{SiO}_2$ , 200–300 or 300–400 mesh; *Qingdao Marine Chemical Factory*). Anal. TLC: silica-gel plates ( $\text{SiO}_2$ ; *Yantai Institute of Chemical Technology*); eluent petroleum ether/acetone 3 : 1; visualization under UV light, and by spraying with 10% aq.  $\text{H}_2\text{SO}_4$  soln., followed by heating. Optical rotations (ORD): *Jasco-P-1020* spectropolarimeter. UV Spectra: *Shimadzu-UV-260* spectrophotometer; in anh. MeOH;  $\lambda_{\text{max}}$  (log  $\epsilon$ ) in nm. IR Spectra: *Avatar-360-ESP* spectrophotometer (*Thermo Nicolet*); KBr pellets;  $\tilde{\nu}$  in  $\text{cm}^{-1}$ .  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR Spectra: *DRX-400* spectrometer; in  $\text{CDCl}_3$ ;  $\delta$  in ppm rel. to  $\text{Me}_4\text{Si}$  as internal standard,  $J$  in Hz. HR-ESI-MS: *Bruker-Apex-70-Tesla* FT-MS apparatus; in  $m/z$ .

*Plant Material.* The aerial parts of *Solidago canadensis* L. were collected in Shanghai, P. R. China, in August 2006. A voucher specimen (Qin-Zeng200601) is deposited with the Herbarium of Materia Medica, School of Pharmacy, the Second Military Medical University, Shanghai, P. R. China.

*Extraction and Isolation.* The air-dried, aerial parts (20 kg) of *S. canadensis* were extracted exhaustively with 80% aq. EtOH at r.t. The EtOH extract was concentrated to yield a semi-solid (1424 g), which was suspended in  $\text{H}_2\text{O}$  (1000 ml), and extracted with AcOEt ( $3 \times 600$  ml). The combined AcOEt phase was concentrated to yield a residue (105 g), part of which (100 g) was subjected to CC (2 kg of  $\text{SiO}_2$ , petroleum ether/AcOEt gradient): *Fractions 1–7*. *Fr. 5* (eluted with petroleum ether/AcOEt 3 : 1) was subjected to repeated CC ( $\text{SiO}_2$ ; petroleum ether/AcOEt 4 : 1), and then to prep. TLC (petroleum ether/AcOEt 7 : 2): **1** (15 mg) and **2** (8 mg). *Fr. 4* (eluted with petroleum ether/AcOEt 5 : 1), was subjected to repeated CC ( $\text{SiO}_2$ , petroleum ether/AcOEt 8 : 1): **3** (6 mg). *Fr. 2* (eluted with petroleum ether/AcOEt 10 : 1) was subjected to repeated CC ( $\text{SiO}_2$ , petroleum ether/AcOEt 12 : 1), and then to prep. TLC (petroleum ether/acetone 20 : 1): **4** (4 mg).

*Solidagocanin A* (= (2*Z*)-3-Methyl-5-[(1*S*,2*R*,4*R*,4*aR*,8*aR*)-1,2,3,4,4*a*,7,8,8*a*-octahydro-1,2,4*a*,5-tetramethyl-4-[(2*E*)-2-methyl-1-oxobut-2-en-1-yl]oxy]naphthalen-1-yl]pent-2-enoic Acid; **1**): White powder.  $[\alpha]_{\text{D}}^{25} = +32$  ( $c = 0.05$ , MeOH). UV (MeOH): 219 (3.52). IR (KBr): 3441, 2967, 1700, 1694, 1636, 1437, 1256, 1156, 1072.  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR: *Tables 1* and *2*. HR-ESI-MS: 403.2835 ( $[M + \text{H}]^+$ ,  $\text{C}_{25}\text{H}_{39}\text{O}_4^+$ ; calc. 403.2849).

*Solidagocanin B* (= (2*Z*)-3-Methyl-5-[(1*S*,2*R*,4*R*,4*aR*,8*aR*)-1,2,3,4,4*a*,7,8,8*a*-octahydro-1,2,4*a*,5-tetramethyl-4-[(2*Z*)-2-methyl-1-oxobut-2-en-1-yl]oxy]naphthalen-1-yl]pent-2-enoic Acid; **2**): White powder.  $[\alpha]_{\text{D}}^{25} = +36$  ( $c = 0.05$ , MeOH). UV (MeOH): 218 (3.86). IR (KBr): 3429, 2967, 1710, 1690, 1639, 1438, 1385, 1232, 1158.  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR: *Tables 1* and *2*. HR-ESI-MS: 403.2832 ( $[M + \text{H}]^+$ ,  $\text{C}_{25}\text{H}_{39}\text{O}_4^+$ ; calc. 403.2849).

### REFERENCES

- [1] L.-P. Christensen, J. Lam, *Phytochemistry* **1991**, *30*, 2453.
- [2] F. Bohlmann, T. Burkhardt, C. Zdero, 'Naturally Occurring Acetylenes', Academic Press, London, 1973.
- [3] F. Seaman, F. Bohlmann, C. Zdero, T.-J. Mabry, 'Diterpenes of Flowering Plants, Compositae (Asteraceae)', Springer, New York, 1990.
- [4] T. Lu, M. A. Menelaou, D. Vargas, F. R. Fronczek, N. H. Fischer, *Phytochemistry* **1993**, *32*, 1483.

- [5] M.-E. Bradette-Hébert, J. Legault, S. Lavoie, A. Pichette, *Chem. Pharm. Bull.* **2008**, *56*, 82.
- [6] X.-Y. Liu, H.-K. Zhu, S.-H. Wu, Y.-L. Chen, F.-Y. Liu, P. Wu, *J. Zhejiang Univ. (Sci. Ed.)* **2007**, *34*, 661.
- [7] W.-H. Ma, Y. Lu, H. Huang, P. Zhou, D.-F. Chen, *Bioorg. Med. Chem. Lett.* **2009**, *19*, 4958.
- [8] C.-M. Sun, R.-F. Toia, *J. Nat. Prod.* **1993**, *56*, 953.
- [9] O. F. Smetanina, A. I. Kalinovskii, Y. V. Khudyakova, N. N. Slinkina, M. V. Pivkin, T. A. Kuznetsova, *Chem. Nat. Compd.* **2007**, *43*, 395.

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